

利用生物法製備幾丁質

簡子婷(5140)

2020/05/13

大綱

一、前言

二、蝦殼之前處理及組成分析

三、多孔性蝦殼之製備及其性質分析

四、利用乳酸菌進行生物法處理及其分析

五、結論

摘要

幾丁質(chitin)為世界上含量最多的含氮多醣，主要存在於節肢動物、甲殼類動物(蝦、蟹等)、昆蟲等的外骨骼和真菌、藻類細胞壁中，幾丁質和其衍生物幾丁聚醣(chitosan)，因具有生物相容性、生物降解性和無毒性等特性，而被廣泛的應用於產業中。傳統製備幾丁質的方法，乃利用強酸、強鹼分別進行脫礦物質及脫蛋白質，具有好的生產效率及純度，但會造成環境危害以及聚合度的下降。本研究為利用友善環境以及綠色製程方法，先經由氣爆膨發或冷凍解凍循環處理，使原料產生多孔性結構，再以微生物發酵去除蝦殼的礦物質(主要為碳酸鈣)和蛋白質，製備出幾丁質。結果顯示經氣爆膨發後的蝦殼，其孔徑為 $1.22 \pm 0.24 \mu\text{m}$ ，而冷凍解凍後的蝦殼孔徑為 $0.63 \pm 0.14 \mu\text{m}$ ，皆大於未經處理蝦殼的 $0.47 \pm 0.14 \mu\text{m}$ 。而後經由三株產酸菌，分別為 *Lactobacillus acidophilus* BCRC 10695、*Lactobacillus plantarum* BCRC 10069、*Lactobacillus helveticus* BCRC 14026 進行微生物發酵，結果顯示在發酵 4 天後，*L. plantarum* BCRC 10069 其脫礦物質率(DM)為最高 92.2%，*L. helveticus* BCRC 14026 脫礦物質率(DM)為 84.8%，*L. acidophilus* BCRC 10695 DM 為 69.0%，其中又以 *L. plantarum* BCRC 10069 在 37°C、180 rpm 環境下發酵蝦殼 4 天的條件為最佳。

參考文獻

- 郭芷良，2017，以物理性方法製備幾丁質奈米纖維，國立台灣海洋大學食品科學系碩士論文，基隆，台灣。
- Castro, R., Guerrero-Legarreta, I., & Bórquez, R. (2018). Chitin extraction from *Allopetrolisthes punctatus* crab using lactic fermentation. *Biotechnology Reports*, 20, e00287.
- Doan, C. T., Tran, T. N., Vo, T. P. K., Nguyen, A. D., & Wang, S.-L. (2019). Chitin extraction from shrimp waste by liquid fermentation using an alkaline protease-producing strain, *Brevibacillus parabrevis*. *International Journal of Biological Macromolecules*, 131, 706-715.
- Dun, Y., Li, Y., Xu, J., Hu, Y., Zhang, C., Liang, Y., & Zhao, S. (2019). Simultaneous fermentation and hydrolysis to extract chitin from crayfish shell waste. *International Journal of Biological Macromolecules*, 123, 420-426.
- Ghorbel-Bellaaj, O., Younes, I., Maâlej, H., Hajji, S., & Nasri, M. (2012). Chitin extraction from shrimp shell waste using *Bacillus bacteria*. *International Journal of Biological Macromolecules*, 51(5), 1196-1201.
- Guo, N., Sun, J., Zhang, Z., & Mao, X. (2019). Recovery of chitin and protein from shrimp head waste by endogenous enzyme autolysis and fermentation. *Journal of Ocean University of China*, 18(3), 719-726.
- Hajji, S., Ghorbel-Bellaaj, O., Younes, I., Jellouli, K., & Nasri, M. (2015). Chitin extraction from crab shells by *Bacillus bacteria*. Biological activities of fermented crab supernatants. *International Journal of Biological Macromolecules*, 79, 167-173.
- Hoke, K., Houška, M., Průchová, J., Gabrovská, D., Vaculová, K., & Paulíčková, I. (2007). Optimisation of puffing naked barley. *Journal of Food Engineering*, 80(4), 1016-1022.
- Jang, M. K., Kong, B. G., Jeong, Y. I., Lee, C. H., & Nah, J. W. (2004). Physicochemical characterization of α -chitin, β -chitin, and γ -chitin separated from natural resources. *Journal of Polymer Science Part A: Polymer Chemistry*, 42(14), 3423-3432.
- Kaur, S., & Dhillon, G. S. (2015). Recent trends in biological extraction of chitin from marine shell wastes: a review. *Critical Reviews in Biotechnology*, 35(1), 44-61.
- Lee, S., & Lee, J. (2009). Effects of oven-drying, roasting, and explosive puffing process on isoflavone distributions in soybeans. *Food Chemistry*, 112(2), 316-320.
- Liu, P., Liu, S., Guo, N., Mao, X., Lin, H., Xue, C., & Wei, D. (2014). Cofermentation of *Bacillus licheniformis* and *Gluconobacter oxydans* for chitin extraction from shrimp waste. *Biochemical Engineering Journal*, 91, 10-15.

- 1 Lu, Y., Sun, Q., She, X., Xia, Y., Liu, Y., Li, J., & Yang, D. (2013). Fabrication and
2 characterisation of α -chitin nanofibers and highly transparent chitin films by pulsed
3 ultrasonication. *Carbohydrate Polymers*, 98(2), 1497-1504.
- 4 Mao, X., Guo, N., Sun, J., & Xue, C. (2017). Comprehensive utilization of shrimp waste
5 based on biotechnological methods: A review. *Journal of Cleaner Production*, 143,
6 814-823.
- 7 Mao, X., Zhang, J., Kan, F., Gao, Y., Lan, J., Zhang, X., Hu, Z., Li, Y., Lin, H. (2013).
8 Antioxidant production and chitin recovery from shrimp head fermentation with
9 *Streptococcus thermophilus*. *Food Science and Biotechnology*, 22(4), 1023-1032.
- 10 Nesvadba, P. (2008). Thermal properties and ice crystal development in frozen foods. In:
11 *Frozen Food Science Technology*. Evans, J. A. (Ed), Blackwell Publish, Oxford, UK.
12 pp 1-25.
- 13 Neves, A. C., Zanette, C., Grade, S. T., Schaffer, J. V., Alves, H. J., & Arantes, M. K. (2017).
14 Optimization of lactic fermentation for extraction of chitin from freshwater shrimp
15 waste. *Acta Scientiarum. Technology*, 39(2), 125-133.
- 16 Rødde, R. H., Einbu, A., & Vårum, K. M. (2008). A seasonal study of the chemical
17 composition and chitin quality of shrimp shells obtained from northern shrimp
18 (*Pandalus borealis*). *Carbohydrate Polymers*, 71(3), 388-393.
- 19 Rinaudo, M. (2006). Chitin and chitosan: properties and applications. *Progress in Polymer*
20 *Science*, 31(7), 603-632.
- 21 Sedaghat, F., Yousefzadi, M., Toiserkani, H., & Najafipour, S. (2016). Chitin from *Penaeus*
22 *merguiensis* via microbial fermentation processing and antioxidant activity.
23 *International Journal of Biological Macromolecules*, 82, 279-283.
- 24 Sedaghat, F., Yousefzadi, M., Toiserkani, H., & Najafipour, S. (2017). Bioconversion of
25 shrimp waste *Penaeus merguiensis* using lactic acid fermentation: An alternative
26 procedure for chemical extraction of chitin and chitosan. *International Journal of*
27 *Biological Macromolecules*, 104, 883-888.
- 28 Sorokulova, I., Krumnow, A., Globa, L., & Vodyanoy, V. (2009). Efficient decomposition of
29 shrimp shell waste using *Bacillus cereus* and *Exiguobacterium acetylicum*. *Journal of*
30 *Industrial Microbiology & Biotechnology*, 36(8), 1123-1126.
- 31 Zhang, H., Yun, S., Song, L., Zhang, Y., & Zhao, Y. (2017). The preparation and
32 characterization of chitin and chitosan under large-scale submerged fermentation level
33 using shrimp by-products as substrate. *International Journal of Biological*
34 *Macromolecules*, 96, 334-339.