

1 利用蛋白質工程及固定化技術改善幾丁聚糖酶之表現量及酵素特性

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5 一、前言

6 二、將 *Aspergillus fumigatus* 來源之幾丁聚糖酶於大腸桿菌胞外表現後以高細
7 胞密度饋料批式發酵生產並改善其表現量

8 三、利用人工建構之雙硫鍵改善 *Bacillus ehimensis* 來源之幾丁聚糖酶之熱穩定
9 性

10 四、將幾丁聚糖酶固定於矽膠上藉以有效水解幾丁聚糖並生成不同尺寸之幾丁
11 寡糖

12 五、結論

13 摘要

14 幾丁聚糖酶 (chitosanase) 為一種可以水解幾丁聚糖 (chitosan) 上 β -1,4-糖
15 苷鍵，進而形成幾丁寡糖 (chitooligosaccharide) 或是葡萄糖胺 (glucosamide) 之
16 酵素，而本篇報告探討利用不同方式改善幾丁聚糖酶之表現量及酵素特性。將
17 *Aspergillus fumigatus* 來源之幾丁聚糖酶基因選殖入大腸桿菌中表現，在 25°C
18 下以 10 g/L 乳糖誘導並添加 1% 甘胺酸 (glycine) 時，酵素活性為 6015 U/mL，
19 此外進行批式發酵 (fed-batch) 兩天，酵素活性更可達至 14000 U/mL。另外，利
20 用定位突變方式增加 *Bacillus ehimensis* 來源之幾丁聚糖酶 EAG1 (chitosanase
21 EAG1) 之雙硫鍵，以提高其熱穩定性。在所有突變型酵素中，以 *Mut5* 表現最
22 好，半衰期由原本 10.5 分鐘增加至 69.3 分鐘，且其催化效率約為原生型酵素
23 之 2 倍。最後，將幾丁聚糖酶固定於矽膠 (silica-gel) 上用以生產幾丁寡糖並進
24 行其特性分析，在 37°C 時，其固定化酵素活性最佳，雖然固定化酵素活性僅有
25 非固定化酵素的 82.3%，但酵素經固定化後能增加其穩定性及連續催化幾丁聚糖
26 能力皆更加良好，因此具有連續生產之優勢。綜合上述結果，不論是以批式發酵、
27 定位突變增加雙硫鍵或是固定化技術，皆能分別有效增加幾丁聚糖酶之表現量、
28 熱穩定性或是重複利用性，使得在生產幾丁寡糖之應用上更加具有優勢。

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